



# NMR spectroscopy of some electrolyte solutions to 1.9 GPa

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## Abstract

Nuclear-magnetic resonance (NMR) spectra of CsCl and LaCl<sub>3</sub> in D<sub>2</sub>O/H<sub>2</sub>O solutions were collected up to pressures of 1.9 GPa using a new NMR probe design that considerably extends the pressure range available for geochemical experiments. The longitudinal-relaxation times ( $T_1$ ) for <sup>2</sup>H compare well with those reported in the previous studies of Lee et al. (1974), who examined lower pressures, and indicate that the probe functions properly. In some experiments, <sup>133</sup>Cs and <sup>1</sup>H NMR spectra could be taken on solutions to pressures well beyond the nominal freezing pressure of D<sub>2</sub>O or H<sub>2</sub>O to form Ice VI (near 0.9 GPa). Freezing to form the high-pressure ice is kinetically slow on an experimental time scale (minutes to hours). The data indicate that the electrolyte concentrations increase the freezing pressure of the solution. This result means that solution NMR spectra can be collected at pressures that are nearly twice the nominal freezing pressure of pure D<sub>2</sub>O or H<sub>2</sub>O. Pulsed-magnetic-field-gradient NMR methods are used to independently measure the self-diffusion coefficient of H<sub>2</sub>O in these solutions, which yields estimates of solution viscosity via the Stokes–Einstein relation. The increased viscosity accounts for the pressure variation of  $T_1$  values as rates of molecular tumbling are affected. Accounting for such changes is essential if NMR spectral line widths are used to infer pressure-enhanced rates of geochemical reactions, such as interconversion of aqueous complexes. © 2016 Elsevier Ltd. All rights reserved.

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## 1. INTRODUCTION

A common geochemical model of electrolyte solutions, the Helgeson-Kirkham-Flowers (HKF) model, describes the partial molar properties of solutes using an expression containing the dielectric constant of water and solute-specific variables derived from fits to experimental data. When this model is coupled to thermodynamic data for minerals, it can be used to estimate solubilities and

speciation information at high pressures and temperatures (Shock and Helgeson, 1988; Tanger and Helgeson, 1988; Shock et al., 1989; Anderson and Crerar, 1993; Shock and Koretsky, 1995; Sverjensky et al., 1997, 2014; Schulte et al., 2001). Recently the HKF model was extended to 6.0 GPa and 1200 °C via molecular-dynamic estimates of the dielectric constant of water (Pan et al., 2013; Sverjensky et al., 2014). These new pressure and temperature limits inspired the design of a nuclear magnetic resonance (NMR) probe for examining solute speciation at higher pressures (Pautler et al., 2014; Ochoa et al., 2015).

There is a rich literature on high-pressure NMR spectroscopy, but this earlier work is generally limited to pressures less than 0.5 GPa (see Ballard et al., 1996), with heroic exceptions (Jonas, 1980; Lang and Lüdemann,

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1993; de Langen and Prins, 1995; Ballard et al., 1998). These earlier designs generally employed milliliter-sized samples. In contrast, the design described in the present paper has a 10–15 microliter sample volume (Fig. 1) and can reach pressures of 2.0 GPa. The early work was directed at detailing solvent motions and relaxation mechanisms (Lee and Jonas, 1971; Lee et al., 1974; Akai and Jonas, 1976; Jonas et al., 1976; Defries and Jonas, 1977; Jonas, 1980; Lamb et al., 1981; Lamb and Jonas, 1981; Lang and Lüdemann, 1993; Ballard et al., 1998). High-pressure NMR was then extended to estimate the activation volumes of homoleptic reactions (Merbach and Vanni, 1977; Asano and Noble, 1978; Ducommun et al., 1979a,b, 1980; Monnerat et al., 1981; Swaddle and Merbach, 1981; Meyer et al., 1982; Hugi-Cleary et al., 1985, 1987; Cossy et al., 1987; Minirale, 1989; Pittet et al., 1990; Takagi et al., 1994; Drljaca et al., 1998a; Swaddle et al., 2005; Dees et al., 2007) with an intent of assigning mechanisms to ligand-exchange reactions (Helm and Merbach, 2005). The work has been reviewed several times (Asano and Noble, 1978; Eldik et al., 1989; Drljaca et al., 1998b). Diamond-anvil technologies, of course, can reach much higher pressures but are limited to nanoliter sample volumes or less, which are too small for studying solutes in water (e.g., Meier et al., 2015).

In this paper, the NMR spectroscopy of a fully dissociated electrolyte (CsCl, LaCl<sub>3</sub>) is studied in order to reproduce the earlier work of Lee et al. (1974) and to demonstrate the utility of the new probe design. Time constants for the longitudinal relaxation ( $T_1$ ) of <sup>2</sup>H were measured on aqueous CsCl in D<sub>2</sub>O and compare well with the original work. In addition, values of  $T_1$  were determined for <sup>133</sup>Cs and <sup>2</sup>H NMR in the solutions and were compared to previous work on LaCl<sub>3</sub> (Lee and Jonas, 1971; Lee et al., 1974; Ochoa et al., 2015). The CsCl and LaCl<sub>3</sub> solutions were chosen because Jonas' group not only measured the  $T_1$  values for these solu-

tions, but also measured viscosities via the rolling-ball method at high pressures. Knowing the viscosity of an experimental solution is important because this controls rates of molecular tumbling and thus affects the widths of NMR spectral peaks.

## 2. EXPERIMENTAL METHODS

All cesium chloride (CsCl) solutions were prepared by dissolving the anhydrous salt into deuterium oxide solvent (D<sub>2</sub>O). Lanthanum chloride (LaCl<sub>3</sub>) solutions were made similarly but in H<sub>2</sub>O with a resistance of 18 MΩ. Solution compositions were verified by coulometric titration for the chloride ion. All pD and pH measurements were made with a combination electrode and calibrated using standard buffers in H<sub>2</sub>O. The pD values were calculated using:  $pD = pH + 0.4$  (Kręzel and Bal, 2004).

The design of the high-pressure NMR probe is described in earlier papers (Pautler et al., 2014; Ochoa et al., 2015) and only the highlights are mentioned here. This high-pressure NMR probe is distinct from previous designs (e.g., Ballard et al., 1996, 1998) because it employs a small solenoid coil made of Berylco-25 wire wrapped around cylindrical capsule of PEEK® (Polyether ether ketone) tubing containing 10–15 μL experimental solution. The cylindrical capsule is sealed at each end (Fig. 1.A) with waterproof epoxy. The solenoid is fabricated from 28 gauge beryllium-copper wire and fed through small holes in the assembly where it is sealed with Stycast® epoxy mixed with a small amount of Al<sub>2</sub>O<sub>3</sub> powder (Fig. 1.B). A ruby attached to a 200 μm fiber-optic cable is also passed through the feedthrough, just below the RF coil, and sealed. Pressure was monitored *in situ* using ruby fluorescence where movement of the R1 peak with pressure (Piermarini et al., 1975; Mao et al., 1986) could be monitored using an Ocean Optics HR4000 UV–vis spectrometer. To ensure that the lowest-pressure measurements were accurate, we confirmed the ruby-fluorescence calibration via a four-wire resistance measurement on manganin wire (Supplemental Information). The correlation is nearly exact (Supplemental Information) but we use the fluorescence estimates of pressure throughout this paper because they can be made *in situ*.

The <sup>133</sup>Cs and <sup>2</sup>H NMR data were acquired using a 300 MHz (7.0 T) Oxford Instruments 78-mm-bore superconducting magnet interfaced to a spectrometer controlled by Tecmag Orion® software. The <sup>2</sup>H  $T_1$  measurements were acquired using a standard inversion-recovery pulse sequence with a calibrated  $\pi/2$  time of 15.8 μs and a relaxation delay of 3 s (applied peak-to-peak radiofrequency voltage of 130 V with a circuit  $Q \sim 11$ ). The <sup>133</sup>Cs  $T_1$  measurements were acquired using a saturation-recovery pulse sequence with a calibrated  $\pi/2$  of 9 μs and a relaxation delay of 100 ms. The probe includes a cooling jacket through which water was circulated at 25(±2) °C from an external water bath. The two degree range corresponded to a diurnal variation. Temperature in the probe was checked with a Type T thermocouple. The experimental range in temperature during an experiment was much smaller at 25 °C (±0.5 °C).

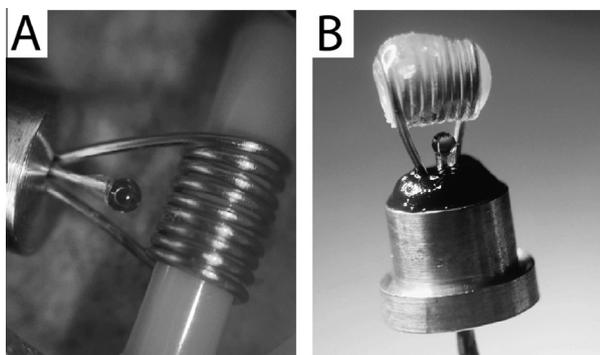


Fig. 1. The microcoil geometry, showing the 1-mm-diameter ruby sphere that is attached to a fiberoptic cable for pressure measurements and calibration. (A): Solenoid and ruby before a solution sample has been placed in the 10–15 μL PEEK® tubing container, and before trimming, with no Stycast® added to seal the beryllium-copper feedthrough. The coil length is 3.4 mm and the coil diameter is 2.7 mm. (B): The solenoid ready for an experiment. The sample is sealed into the tubing and the assembly is sealed with Stycast® mixed with Al<sub>2</sub>O<sub>3</sub> powder.

The self-diffusion coefficients of H<sub>2</sub>O in aqueous solutions were measured using <sup>1</sup>H NMR on a permanent-magnet-based Aspect Imaging MR-100 instrument at 43.7 MHz (1.0 T) that is interfaced to a Tecmag Apollo Spectrometer. A standard pulsed-gradient spin-echo (PGSE) pulse sequence was used with a field-gradient pulse length of  $\delta = 10$  ms and a gradient-pulse spacing of  $\Delta = 100$  ms (Stilbs, 1987; Antalek, 2002). The  $\pi/2$  pulse time was calibrated to 15  $\mu$ s for all solutions. Parameters were extracted from the measured data with a Matlab code by fitting the signal intensity to an exponential decay (Stejskal and Tanner, 1965; Tanner and Stejskal, 1968). Temperature in the bore of this magnet was also monitored via a type-T thermocouple.

### 3. SOLUTION NMR

Rapid tumbling of small molecules in a strong magnetic field causes the longitudinal and transverse relaxation times to become equivalent ( $T_1 = T_2$ ). In this “extreme narrowing” limit, the longitudinal relaxation rates of quadrupolar nuclei ( $I > 0$ ), such as <sup>2</sup>H and <sup>133</sup>Cs can be determined from (Harris, 1986):

$$\frac{1}{T_{1Q}} = \frac{1}{T_{2Q}} = \frac{3}{10} \pi^2 \frac{2I + 3}{I^2(2I - 1)} \langle \chi^2 \rangle \tau_c \quad (1)$$

where  $I$  is the spin of the nucleus,  $\tau_c$  is the reorientation time of the molecule, and  $\langle \chi^2 \rangle$  is the mean square of the zero-average quadrupolar-coupling constant.

The reorientation time of the molecule,  $\tau_c$ , depends upon solution viscosity and can be described using the Stokes–Einstein equation as:

$$\tau_c = \frac{\eta \kappa V_m}{kT} \quad (2)$$

where  $\eta$  is macroscopic viscosity,  $V_m$  is hydrodynamic volume, and  $\kappa$  is a theoretical constant that describes the ratio of intermolecular torques on the solute molecule to the intermolecular forces of the solvent molecules (McClung and Kivelson, 1968; Kivelson et al., 1970). The value for  $\kappa$  is typically set to unity, so that  $V_m$  can be treated as the effective hydrodynamic volume. In these experiments, that volume would be D<sub>2</sub>O, H<sub>2</sub>O, or a single Cs<sup>+</sup> or La<sup>3+</sup> ion in solution.

The solution viscosity increases with pressure, which originally motivated (Lee et al., 1974) to directly measure viscosity in the high-pressure solutions. Here estimates in the pressure-variation of viscosity can be determined via measurements of the diffusion coefficient of H<sub>2</sub>O using the alternative expression:

$$D = \frac{kT}{6\pi\eta r} \quad (3)$$

where  $D$  is the self-diffusion coefficient of molecules in the solvent,  $r$  is the hydrodynamic radius of the molecule and all other variables retain their usual definitions (Edward, 1970). In evaluating Eq. (3), a hydrodynamic radius for water of 1.379 Å is used (Woessner, 1964; Rahman and Stillinger, 1971) and assumed to be independent of pressure.

The diffusion coefficient of H<sub>2</sub>O was measured via a variation of a method employed by Akai and Jonas (1976) that is well developed for ambient pressure (Stejskal and Tanner, 1965; Tanner and Stejskal, 1968). In the PGSE experiment, the apparent signal-decay time constant is measured with a spin-echo experiment as function of the strength of the applied magnetic-field gradient pulses. The Akai and Jonas (1976) method employed a static gradient. For the work reported here, the PGSE pulse sequence is a standard  $\pi/2$ -pulse followed by a  $\pi$ -pulse. Following the first  $\pi/2$ -pulse, an initial magnetic-field gradient pulse is applied with length  $\delta$ . Subsequently, at a time  $\Delta$  later than the initial gradient, a second gradient pulse with an identical length  $\delta$  is applied. The apparent diffusion coefficient is determined by fitting the measured intensity data to the following equation (Stejskal and Tanner, 1965; Tanner and Stejskal, 1968):

$$S = S_0 \exp \left[ -\gamma^2 G^2 \delta^2 \left( \Delta - \frac{\delta}{3} \right) D \right] \quad (4)$$

where  $S_0$  is signal intensity in the absence of a gradient,  $\gamma$  is the gyromagnetic ratio,  $G$  is the strength of the gradient pulse in Tesla/m, and  $D$  is the apparent diffusion coefficient. During the field gradient pulse, the signal intensity,  $S$ , decreases exponentially as a function of time and as a function of  $G$ . These intensity data are then fit to Eq. (4) in order to calculate  $D$ , the apparent diffusion coefficient, with uncertainties established as the precision of four trials. This value is ‘apparent’ because the actual magnetic-field gradients affecting the microcoil sample are not known with certainty, only the field gradients produced by the spectrometer. Conventionally, such geometry-specific effects are eliminated by using a well-accepted standard to scale the measurements. An accepted value for the diffusion coefficient of H<sub>2</sub>O at 20 °C is  $2.025 \times 10^{-9}$  m<sup>2</sup>/s (Holz et al., 2000). This value is used to scale the apparent measurements, which are within a factor of two of this number at ambient pressures. An assumption is made that the scaling does not vary with pressure, which is reasonable since controls of the magnetic-field gradients are exterior to the probe. The coil and sample are also not considerably deformed by the hydrostatic pressure.

## 4. RESULTS

### 4.1. <sup>2</sup>H and <sup>133</sup>Cs NMR relaxation rates in CsCl solutions compared to LaCl<sub>3</sub> solutions

The  $T_1$  values for <sup>2</sup>H in various solutions at 25 °C are shown in Fig. 2.A. The values for pure D<sub>2</sub>O at 25 °C (this paper) and at 10 °C and 30 °C from Jonas’ group (Lee et al., 1974) are shown in Fig. 2.B. The results using the microcoil probe are bracketed by the previous work at near temperatures (Fig. 2.B). The similarity between the  $T_1$  values reported here and those reported previously indicates that the microcoil sample is not being heated by repeated pulsing and that the NMR probe design is trustworthy. The maximum in  $T_1$  values with pressure has generally been interpreted to indicate pressure-enhanced structuring of the solvent. The microcoil design has much less precise control

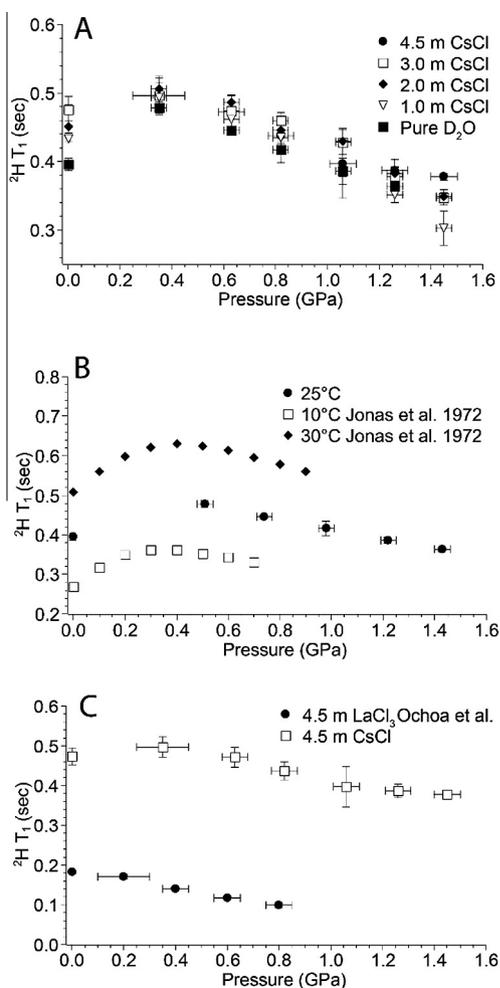


Fig. 2. (A):  $^2\text{H}$   $T_1$  values as a function of pressure for various solutions at  $25(\pm 0.5)$  °C. (B):  $^2\text{H}$   $T_1$  data values as a function of pressure for pure  $\text{D}_2\text{O}$  compared with previous data (Lee and Jonas, 1972). (C): Values of  $^2\text{H}$   $T_1$  for CsCl and  $\text{LaCl}_3$  solutions (Ochoa et al., 2015) at  $25(\pm 0.5)$  °C. Errors assigned to pressure ( $\pm 30$  MPa) was determined by propagating a  $\pm 0.01$  nm estimated uncertainty in the R1 fluorescence peak position through the equations relating the ruby fluorescence shift to pressure (Dewaele et al., 2008). Uncertainties in the  $T_1$  values correspond to the estimated standard deviation of triplicate measurements.

over pressure at  $P < 0.4$  GPa than the large-volume design of Lee et al. (1974), but it can recapture the characteristic maxima in  $^2\text{H}$   $T_1$  values observed for all CsCl solutions. Lee et al. (1974) report the maximum at about 0.3–0.5 GPa. Comparison of  $^2\text{H}$   $T_1$  values in a 4.5 m CsCl and  $\text{LaCl}_3$  solutions indicate that the steady decline in  $T_1$  values with pressure is similar (Fig. 2.C). Beyond the 0.5 GPa maxima, the  $T_1$  values for all  $\text{D}_2\text{O}$  solutions decrease uniformly (Fig. 2) and this decrease has been interpreted as resulting from an increase in solution viscosity (Lee et al., 1974).

The  $^{133}\text{Cs}$   $T_1$  values of CsCl solutions (Fig. 3) also decrease uniformly with pressure and the rate of decrease is nearly, but not completely, independent of solution composition. These results contrast with previous work on

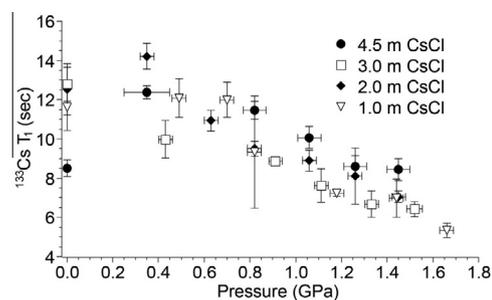


Fig. 3.  $T_1$  values for  $^{133}\text{Cs}$  for various CsCl solutions in  $\text{D}_2\text{O}$  at  $25(\pm 0.5)$  °C. Errors assigned to pressure ( $\pm 30$  MPa) for 1.0 m, 2.0 m, and 3.0 m solutions, were determined by propagating a  $\pm 0.01$  nm estimated uncertainty in the R1 fluorescence peak position through the equations relating the ruby fluorescence shift to pressure (Dewaele et al., 2008). Uncertainties for the 4.5 m CsCl solution ( $\pm 100$  MPa) were determined using a standard curve relating hydraulic press force to pressure measured in separate samples from ruby fluorescence (Ochoa et al., 2015). Uncertainties in the  $T_1$  values correspond to a single estimated standard deviation of triplicate measurements.

$\text{LaCl}_3$  solutions where the  $^{139}\text{La}$   $T_1$  values vary considerably with electrolyte concentrations at ambient pressures (Ochoa et al., 2015). For example, at ambient pressures, an increase of  $\text{LaCl}_3$  concentration from 1.0 m to 4.5 m is accompanied by a large decrease in the  $T_1$  values from  $1.11(\pm 0.02)$  ms to  $0.18(\pm 0.09)$  ms (Ochoa et al., 2015). In contrast,  $T_1$  values for  $^{133}\text{Cs}$  decrease only from  $11.6(\pm 1.2)$  s to  $8.5(\pm 0.4)$  s over a similar range in concentration. The difference is also manifested in the properties of the solvent. For example, the  $T_1$  values for  $^2\text{H}$  in 1.0 m and 4.5 m  $\text{LaCl}_3$  are  $0.27(\pm 0.02)$  and  $0.18(\pm 0.005)$  s, respectively. This difference is much larger than in similar CsCl solutions, where the  $^2\text{H}$   $T_1$  values are  $0.43(\pm 0.04)$  s and  $0.47(\pm 0.02)$  s for 1.0 m and 4.5 m CsCl solutions, respectively.

#### 4.2. Diffusion coefficients for $\text{H}_2\text{O}$ at pressure

As was originally concluded by the Jonas group, pressure variations in  $T_1$  values reflect changes in viscosity of the solution. As described above, the viscosity can be estimated from measurement of an apparent diffusion coefficient using the PGSE pulse sequence. The apparent diffusion coefficients for  $\text{H}_2\text{O}$  in both pure water and in a 1.0 m CsCl solution measured here are shown in Fig. 4.A. Note that the D values for both solutions decrease uniformly with pressure and that these trends match well the pressure variation of  $T_1$  values shown in Figs. 2 and 3. Solutions viscosities were calculated via Eq. (3) and are reported in Table 1.

If changes in viscosity alone caused the measured variation in  $T_1$  values with pressure, then the ratio of  $D/T_1$  would be independent of pressure. Such a ratio is shown as Fig. 4.B. As one can see, the ratios for all conditions lie within 10 % of each other and are independent of pressure, within experimental error.

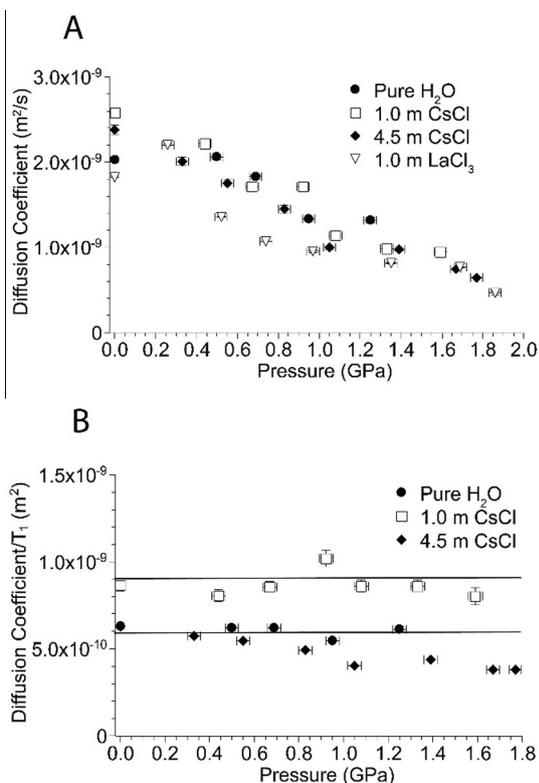


Fig. 4. (A): Self-diffusion coefficients for H<sub>2</sub>O in water and aqueous solutions of 1.0 m CsCl, 4.5 m CsCl, or 1.0 m LaCl<sub>3</sub> at 20 (±0.5) °C. (B): The diffusion coefficients divided by the <sup>1</sup>H T<sub>1</sub> measurements for pure H<sub>2</sub>O and for aqueous 1.0 m CsCl. Uncertainties from pressure were determined by propagating a ±0.01 nm (±30 MPa) estimated uncertainty in the peak position through the equations relating the R1 fluorescence of the ruby to pressure (Dewaele et al., 2008). Error estimates assigned to diffusion coefficient values are the estimated standard deviation from four repeated PGSE trials. The lines are merely to guide the eye and are not regressions.

### 4.3. Evidence for suppressed freezing

Pressure was applied to all CsCl solutions until they froze, which was inferred from the disappearance of the <sup>133</sup>Cs NMR peak, or when the <sup>2</sup>H linewidth increased beyond the bandwidth of the detector. When pressure was subsequently relieved in order to return the sample to ambient conditions, the NMR spectra were reacquired and yielded the T<sub>1</sub> value expected for the homogeneous solution, as one would expect for reversible freezing. The estimated pressures when each solution froze are reported in Fig. 5, with respect to various phase boundaries. Ice-V and Ice-VI phase boundaries were reproduced from the results of Wagner et al. (1994). The phase boundary for metastable ice was reproduced from Whittaker et al. (1998) and is labeled in the figure as ‘new ice’. Lines in Fig. 5 also identify the solid–liquid phase boundary in the presence of different NaCl concentrations in H<sub>2</sub>O (Journaux et al., 2013). As one can see, the experimental freezing points exceed by almost a factor of two the pressures expected from the phase boundary between liquid

Table 1

Diffusion coefficients and calculated viscosities for pure water and for 1.0 m CsCl solution at 20(±0.5) °C. Diffusion coefficients in this table are estimated by dividing the measured value at ambient pressure by an accepted diffusion coefficient value for water at 20 °C of 2.025 × 10<sup>-9</sup> m<sup>2</sup>/s (Holz et al., 2000). Uncertainties are reported as an estimated standard deviation. Uncertainties in the pressure estimates are ±0.03 GPa.

Pressure (GPa)	Diffusion (m <sup>2</sup> /s)	Viscosity (cP)
<i>Pure H<sub>2</sub>O</i>		
1.01 × 10 <sup>-4</sup>	2.03 × 10 <sup>-9</sup> ± 2.8 × 10 <sup>-11</sup>	0.77 ± 0.01
0.50	2.06 × 10 <sup>-9</sup> ± 2.8 × 10 <sup>-11</sup>	0.75 ± 0.01
0.69	1.83 × 10 <sup>-9</sup> ± 2.05 × 10 <sup>-11</sup>	0.85 ± 0.01
0.95	1.33 × 10 <sup>-9</sup> ± 1.1 × 10 <sup>-11</sup>	1.17 ± 0.01
1.25	1.32 × 10 <sup>-9</sup> ± 3.0 × 10 <sup>-11</sup>	1.18 ± 0.03
<i>1.0 m CsCl in H<sub>2</sub>O</i>		
1.01 × 10 <sup>-4</sup>	2.58 × 10 <sup>-9</sup> ± 2.3 × 10 <sup>-11</sup>	0.60 ± 0.005
0.44	2.22 × 10 <sup>-9</sup> ± 2.6 × 10 <sup>-11</sup>	0.70 ± 0.01
0.67	1.71 × 10 <sup>-9</sup> ± 1.9 × 10 <sup>-11</sup>	0.91 ± 0.01
0.92	1.71 × 10 <sup>-9</sup> ± 1.6 × 10 <sup>-11</sup>	0.91 ± 0.01
1.08	1.14 × 10 <sup>-9</sup> ± 1.1 × 10 <sup>-11</sup>	1.37 ± 0.01
1.33	9.87 × 10 <sup>-10</sup> ± 9.9 × 10 <sup>-12</sup>	1.58 ± 0.03
1.59	9.49 × 10 <sup>-10</sup> ± 1.4 × 10 <sup>-11</sup>	1.64 ± 0.02
<i>4.5 m CsCl in H<sub>2</sub>O</i>		
1.01 × 10 <sup>-4</sup>	2.38 × 10 <sup>-9</sup> ± 5.1 × 10 <sup>-11</sup>	0.65 ± 0.01
0.33	2.01 × 10 <sup>-9</sup> ± 4.8 × 10 <sup>-11</sup>	0.78 ± 0.02
0.55	1.75 × 10 <sup>-9</sup> ± 4.8 × 10 <sup>-11</sup>	0.89 ± 0.02
0.83	1.45 × 10 <sup>-9</sup> ± 4.8 × 10 <sup>-11</sup>	1.07 ± 0.04
1.05	9.99 × 10 <sup>-10</sup> ± 1.95 × 10 <sup>-11</sup>	1.56 ± 0.03
1.39	9.79 × 10 <sup>-10</sup> ± 1.95 × 10 <sup>-11</sup>	1.59 ± 0.03
1.67	7.44 × 10 <sup>-10</sup> ± 1.5 × 10 <sup>-11</sup>	2.09 ± 0.04
1.77	6.47 × 10 <sup>-10</sup> ± 1.1 × 10 <sup>-11</sup>	2.40 ± 0.04
<i>1.0 m LaCl<sub>3</sub> in H<sub>2</sub>O</i>		
1.01 × 10 <sup>-4</sup>	1.83 × 10 <sup>-9</sup> ± 5.2 × 10 <sup>-12</sup>	0.85 ± 0.01
0.26	2.20 × 10 <sup>-9</sup> ± 1.4 × 10 <sup>-11</sup>	0.71 ± 0.01
0.52	1.36 × 10 <sup>-9</sup> ± 5.1 × 10 <sup>-12</sup>	1.15 ± 0.01
0.74	1.07 × 10 <sup>-9</sup> ± 2.1 × 10 <sup>-12</sup>	1.45 ± 0.01
0.97	9.57 × 10 <sup>-10</sup> ± 8.2 × 10 <sup>-12</sup>	1.63 ± 0.01
1.35	8.14 × 10 <sup>-10</sup> ± 1.0 × 10 <sup>-11</sup>	1.91 ± 0.02
1.69	7.66 × 10 <sup>-10</sup> ± 4.2 × 10 <sup>-12</sup>	2.03 ± 0.01
1.86	4.67 × 10 <sup>-10</sup> ± 1.8 × 10 <sup>-11</sup>	3.34 ± 0.13

water and Ice VI of ~0.9 GPa. The points plotted on Fig. 5, however, correspond to a crude estimate of the actual freezing pressure – we cannot identify frozen samples in our NMR probe. We can only report the last pressure where the NMR spectra exhibited liquid-like behavior, where the T<sub>1</sub> values, for example, could be measured, and the jumps in pressure were sufficiently large that the plotted data do not indicate phase boundaries.

## 5. DISCUSSION

### 5.1. Relaxation in fully dissociated electrolytes

The differences in T<sub>1</sub> relaxation values at ambient conditions for solutions of various CsCl and LaCl<sub>3</sub> concentrations are expected since these electrolytes affect the structure of water differently. Electrolytes are classified as ‘structure-breaking’ or ‘structure-making’ according to their effects on NMR relaxation and the structure of water

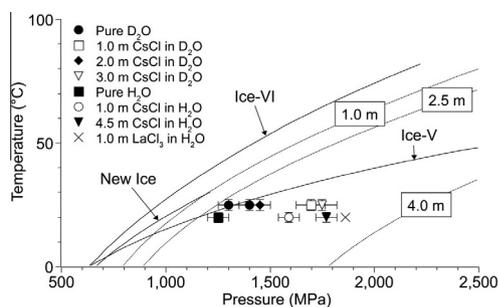


Fig. 5. Estimated minimum freezing pressures of the experimental solutions relative to various phase boundaries for water. These pressures are inferred from the disappearance of liquid-like NMR behavior. Curves for Ice-V and Ice-VI were constructed from Wagner et al. (1994). The curve for metastable ice, labeled ‘New Ice’ was reproduced from Whittaker et al. (1998). The curves describing the salt-affected freezing of NaCl solutions are shown as various lines labeled: ‘4.0 m’, ‘2.5 m’ and ‘1.0 m’ (from Journaux et al., 2013). A sample of 1.0 m LaCl<sub>3</sub> showed liquid-like NMR spectra all the way to 1.86 GPa and the final pressure is shown as an ‘x’.

(Cox and Wolfenden, 1934; Hribar et al., 2002; Marcus, 2009). Structure-breaking ions, like Cs<sup>+</sup>, allow solvent molecules to rotate more freely when added to pure water and structure-making ions, like La<sup>3+</sup>, restrict the rotation of solvent molecules by binding tightly to the bulk solvent lattice. Thus, addition of CsCl or LaCl<sub>3</sub> salts to water has a different effect on the  $T_1$  values of <sup>133</sup>Cs, <sup>139</sup>La and <sup>2</sup>H. Although LaCl<sub>3</sub> solutions exhibit a range of <sup>139</sup>La  $T_1$  values for different electrolyte concentrations at ambient conditions, the <sup>139</sup>La  $T_1$  values decrease and become nearly equal for different LaCl<sub>3</sub> concentrations at  $P > 1.5$  GPa (Ochoa et al., 2015). Pressure eliminates the differences in  $T_1$  values for different solutions.

## 5.2. Viscosity of aqueous solutions at pressure and NMR spectroscopy

The decrease in <sup>2</sup>H and <sup>133</sup>Cs  $T_1$  values to 1.7 GPa are largely attributable to increases in viscosity, leading to corresponding increases in the reorientation times and shorter  $T_1$  values. This result is important. The broadening with

pressure of these signals can, under ideal circumstances, be interpreted to indicate pressure-induced changes in reaction rates and thus activation volumes. Although high-pressure NMR studies of reaction kinetics are usually over a narrow range of pressure (0.1–0.35 GPa) where viscosity-caused line broadening may be minimal, the contribution of viscosity will be increasingly important as high pressures are reached. As one can see in Tables 1 and 2, the increases in solution viscosities with pressure are monotonic and can probably be predicted from a simple regression. This agreement is important because the data in Table 2 were measured by Jonas’ group using a completely different method. They employed a rolling sphere to estimate the viscosity from optical measurements. Our data using the NMR method not only compare well with these independent methods, but also show approximately the same variation with pressure.

## 5.3. Suppressed freezing and metastability

There are two causes of the apparent overpressurization of these solutions. First, the NaCl electrolyte concentration has been shown by Journaux et al. (2013) to suppress the freezing of H<sub>2</sub>O at pressure. Although our electrolyte is CsCl in D<sub>2</sub>O and not NaCl in H<sub>2</sub>O, it is reasonable to expect a similar suppression of the freezing point in a Raoult’s Law sense. The differences between the freezing point of pure D<sub>2</sub>O and H<sub>2</sub>O at high pressures are very small (Brown and Whalley, 1966; Pistorius et al., 1968).

Secondly, freezing is slow in these experiments. An example is shown in Fig. 6 for the 3.0 m CsCl solution. The figure demonstrates solution freezing at 1.75 GPa after 15 min, as gauged by the slow disappearance of the NMR signal. As mentioned above, temperature estimates seem to be accurate and indicate that the sample is not heated by the radio frequency pulse in the saturation-recovery pulse sequence. Similarly, there is no evidence for a pressure gradient, such as would be established if the pressure medium froze (Koyama-Nakazawa et al., 2007). Similar over-pressurization of the samples is observed when either fluorocarbon liquid or Daphne 7373 oil is used as the pressure medium. Both of these liquids are expected to remain liquid at these conditions. The simplest explanation for the relatively high freezing pressures is that higher pressures

Table 2

Viscosity measurements via the rolling-ball method as a function of pressure for pure H<sub>2</sub>O and 4.5 m CsCl in D<sub>2</sub>O at 10 °C and 30 °C (Jonas et al., 1976). Note that these values are about the same size as those reported in Table 1 that were measured independently via NMR methods, and that the change is similar with pressure.

Pressure (GPa)	Viscosity (cP) pure H <sub>2</sub> O at 10 °C	Viscosity (cP) pure H <sub>2</sub> O at 30 °C	Viscosity (cP) 4.5 m CsCl in D <sub>2</sub> O at 30 °C
$1.013 \times 10^{-4}$	1.31	0.801	0.966
0.1	1.26	0.811	1.001
0.2	1.29	0.846	1.059
0.3	1.37	0.899	1.156
0.4	1.47	0.963	1.252
0.5	1.60	1.04	
0.6	1.76	1.13	
0.7	1.94	1.23	
0.8		1.34	
0.9		1.45	

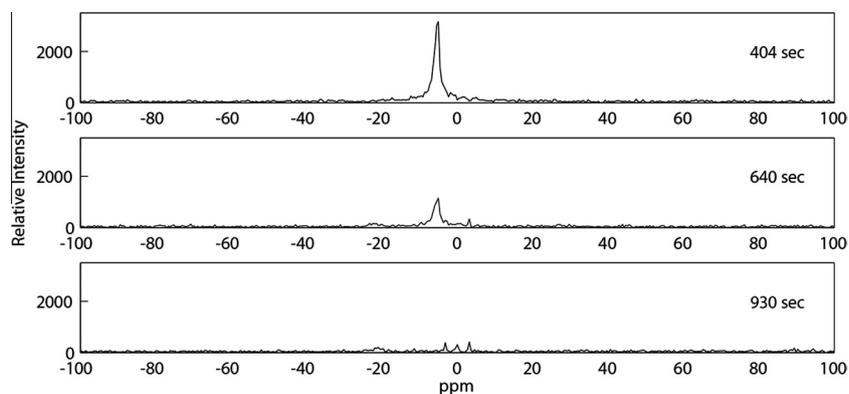


Fig. 6. Sequential  $^{133}\text{Cs}$  NMR spectra for 3.0 m CsCl at 1.75 GPa. The signal loss after 930 s indicates freezing. The remaining small peaks seen at 930 s are noise.

can be reached via the combined effects of freezing-point depression by the electrolyte and solution metastability.

## 6. CONCLUSIONS AND IMPLICATIONS FOR GEOCHEMISTRY

There are several important geochemical results from this study. Foremost is that the NMR probe design allows measurements on solutions to 2.0 GPa. The  $T_1$  relaxation times for  $^2\text{H}$ ,  $^{133}\text{Cs}$ , and  $^{139}\text{La}$  in CsCl,  $\text{LaCl}_3$ , and pure  $\text{D}_2\text{O}$  solutions are reproducible and match well with the earlier work at similar pressures, indicating that the probe is effective. Although this study only deals with fully dissociated electrolytes, the probe can resolve signals from various aqueous complexes (Pautler et al., 2014), yielding equilibrium constants and kinetic information about chemical reaction rates. Thermodynamic data about speciation is codified as reaction volumes and compressibilities (Asano and Noble, 1978; Eldik et al., 1989; Drljaca et al., 1998a), and pressure is particularly useful for understanding the role of solvation or changes in coordination in aqueous reactions (Helm and Merbach, 2005; Swaddle et al., 2005), including isomerization of large molecules (e.g., Johnson et al., 2013).

There is a rich chemistry to explore. As mentioned in the *Introduction*, the range of geochemical models for solution thermodynamics was recently extended via molecular-dynamic estimates of the dielectric properties of water to 6.0 GPa and 1200 °C (Pan et al., 2013; Sverjensky et al., 2014). The advance allowed Sverjensky and Huang (2015) to discover a pH-driven pathway for forming diamonds at 5.0 GPa. These pressures, however, are well beyond the current capabilities of conventional hydrothermal solution spectroscopies, yet through judicious choice of nonmagnetic, high-strength alloys (e.g., Uwatoko et al., 2002), it should not be difficult to extend the 2.0 GPa pressure range of the probe described above to 3.5–4.0 GPa.

The influence of electrolytes on freezing has obvious implications for the transport of liquid water to depths in the Earth, or in salty exoplanets, a point understood by Journaux et al. (2013). Although the freezing points of CsCl and  $\text{LaCl}_3$  aqueous solutions have not yet been evaluated as a function of pressure, it is reasonable to expect a similar

effect to that which is found for NaCl solutions (Journaux et al., 2013). Furthermore, and most interestingly, the crystallization of high-pressure ices from these solutions at pressure is sufficiently slow that NMR spectra can be acquired. We, of course, know nothing of the phases that precipitate when we lose liquid-like NMR signals from these solutions, but these questions await further experimentation.

Finally, transport properties of molecules can be measured easily at pressure, such as the diffusion coefficients shown in Fig. 5. These estimates were previously possible but extraordinarily difficult since the velocities of rolling spheres must be measured at pressure (Abramson, 2007). Yet knowing the viscosity of the solution is essential for using NMR information to infer reaction rates. The activation volumes for the interconversion reactions are often inferred from broadening of the NMR signals as a function of pressure (e.g., Johnson et al., 2013). At the high pressures of this study, contributions to the NMR linewidth from increased solvent viscosity are appreciable, but can be estimated directly and easily (Fig. 5).

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## APPENDIX A. SUPPLEMENTARY DATA

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.gca.2016.08.013>.

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